



Ministry of Agriculture, Livestock and Irrigation

Department of Agriculture

Horticulture and Plant Biotechnology Division



Efficient Tissue Culture Protocol for Propagation of Elephant Foot Yam (*Amorphopallus sp.*)

Presented by

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Introduction

- Tropical tuber crop of south east Asian origin

Traditional

- Vegetable, low calories noodles and artificial meats
- Plant starch (good viscosity and stability) - food industry (**Moorthy *et al.*, 1994**)



Medicinal

- **Starch (11-28%)**, sugar (0.7-1.7%), **protein (0.8-2.60%)**, fat (0.07-0.40%) mean energy value (236-566.70KJ/100g), potassium (327.83 mg/100 g), phosphorus (166.91mg/100 g), calcium (161.08 mg/100 g) and iron (3.43mg/100 g)



(**Anuradha Singh, 2013**)

- High glucomannan content in subterranean corms - reduce the serum cholesterol level (**Cescutti *et al.* 2002 and Ghani 2003**)
- Treatment of tumors, elephantiasis, inflammations, cough, asthma, vomiting, flatulence, colic, dyspepsia, constipation, fatigue and anemia (**Nair, 1993**)

Elephant Foot Yam Production in Southern Chin State

- Major cash crop for Chin farmers
- Produce 100% of households
- Chinese market for 85% of all Myanmar EFY chips
- EFY chips - 5000 MMK/viss
- Japan has permitted 240 metric tonnes of EFY imports from Myanmar
Cambodia, Laos and Sri Lanka (white, yellow and red)
- Yellow and red EFYs are popular export items to China and Japan



Conventional Propagation Methods of EFY



True Seed

- One seed requires about five years to develop into a lager tuber
- Rarely kept or sown



Leaf bulbils

- Collect bulbils at the end of the wet season when the leaf has fallen to the ground
- First-year leaves may produce as few as six smallish bulbils
- After three years, a mature leaf may produce more than 20 bulbils and most favorable propagation material



Tuber

- Small tubers may be re-planted and harvested after one or two years
- Cut one large tuber into vertical wedges & Each wedge may develop into a harvestable tuber after one or two years
- Wedges will rot on the cut surface and fail to grow , Dry and dress the cut with fire ash to reduce the risk of failure

Major Production Constraints

- High incidence of **mosaic disease**
- Non availability of **quality planting materials**
- Cultivated immature yams from wild collection and to reach a suitable size of harvesting for **long time**
- Lack of sufficient seed material of **uniform size and dormancy**



Mosaic virus disease



Various size of tubers

Objectives

- To find out suitable explant source for *in vitro* propagation
- To develop an indirect regeneration protocol for disease free and high quality numerous planting material of EFY within a short period
- To increase agricultural production and income of growers and producers



Materials and Methods

- Sources - Chin state (Kanpetlet)
- Duration - (2019 November - 2020 November)
- Project area - **Plant Biotechnology Center, Palae Myothit, Mingaladon, Yangon**
- Explant types - **leaf, petiole, leaf bulbil, corm bud and floret**
(50 for each explants)



Sterilization of Explants

- Running tap water (30 min)
 - Liquid detergent (15 min) & 0.05% (w/v) fungicide (15 min)
 - 70% (w/v) Ethanol (1 min)
 - 7% or 10% (w/v) NaOCl for 5 min (2 times)
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- Wash 5-7 times with distilled water for each stage
 - Dipping in absolute ethanol and flaming the unopened leaves covered by cataphyll



Five stages for *in vitro* propagation

Initiation stage (establishment of different types of explants)



Callus induction (Modified MS + BAP + NAA + 2,4-D)



Shoot regeneration (Modified MS + BAP + Citric acid + Adenine sulphate)



Root formation (MS, $\frac{1}{2}$ MS, $\frac{1}{4}$ MS + NAA)



Hardening stage

- Modified MS media - half concentration of NH_4NO_3 and KNO_3 (Srinidhi *et al.*, 2008)
- All the media were adjusted to pH 5.8, solidified with 0.7% agar and sterilized by autoclaving at 121°C for 20 min

Statistical analysis

❖ The frequency of callus induction, regeneration and root formation was calculated as follows:

Callus induction frequency (%)	= number of explant producing callus / number of explant plated	× 100
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Regeneration frequency (%)	= number of plants recovered / number of callus plated	× 100
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Root formation frequency (%)	= number of plants recovered / number of callus plated	× 100
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- Factorial complete randomized design (5 treatments × 3 replications)
- ANOVA (Analysis of Variance) & 5% level with (LSD) using Statistix 8.0 software
- Two factors - **Explant types** (leaf, petiole, leaf bulbil, corm bud and floret)
 - **Media combination**

Media Combination

Shoot Regeneration				
No.	Media Combination	BAP (mg/l)	Citric Acid (mg/l)	Adenine Sulphate (g/l)
1.	MSYS1	1	1	0.05
2.	MSYS2	2	1	0.05
3.	MSYS3	3	1	0.05
4.	MSYS4	4	1	0.05
5.	MSYS5	5	1	0.05

Root Formation		
No	Media Combination	
1.	MSYR1	Full MS
2.	MSYR2	Full MS+ NAA (1mg/l)
3.	MSYR3	½ MS+ NAA (1 mg/l)
4.	MSYR4	¼ MS + NAA (1 mg/l)

Results and Discussion

Callus Induction

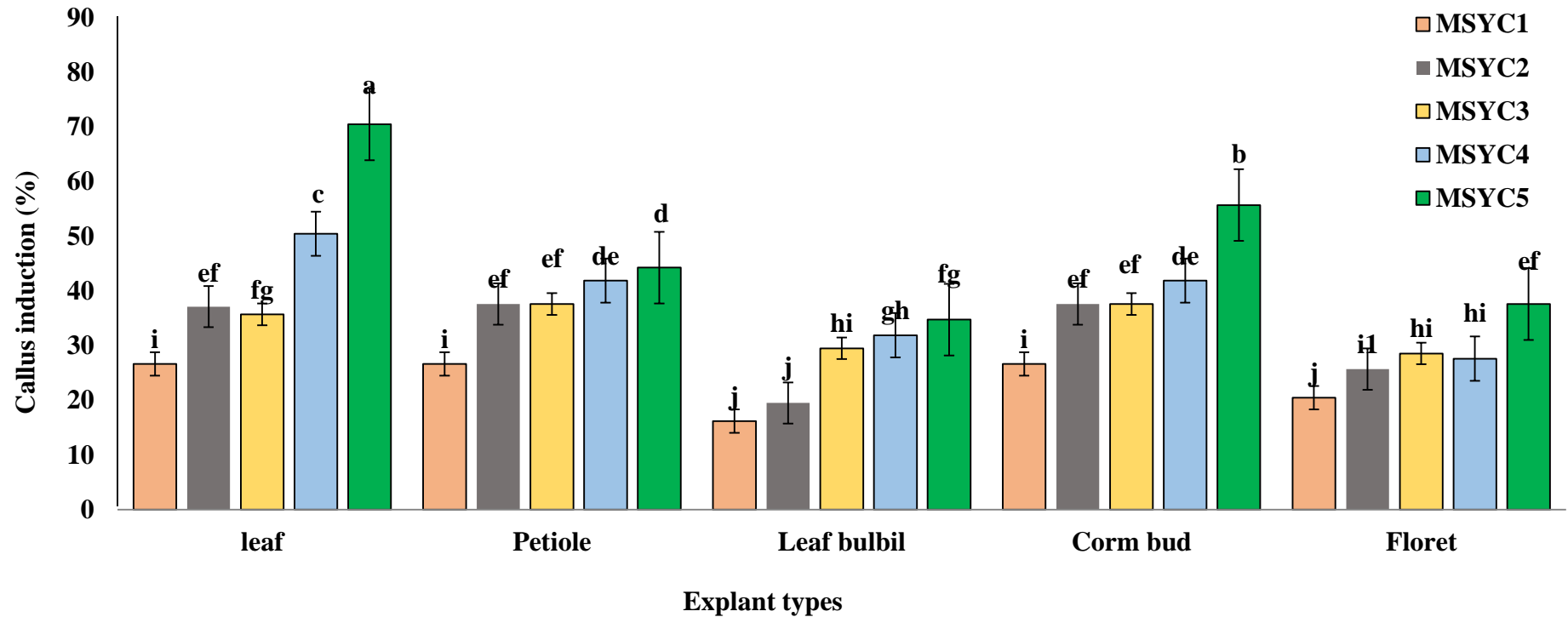


Figure 1. Effect of PGRs on callusing response in different explants of EFY

***** a-j within a column having the same letters are not significantly different at 5% level**

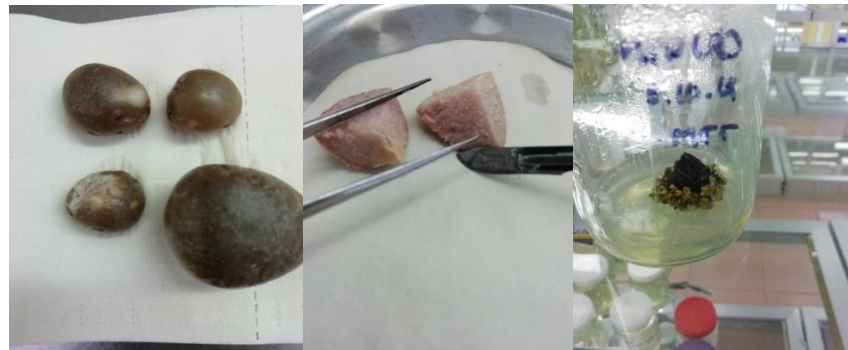
- 2,4-D is one of the most effective auxin for callus induction in sweet potato.
(Oggema, J.N, 2017)
- A combination of 3 PGRs (BAP, NAA and 2,4-D) increase callusing % in petiole and leaf explants of EFY (Kamala and Makesh kumar, 2014)



(a) Young leaf



(b) Petiole



(c) Leaf bulbil



(d) Corm bud



(e) Floret

Figure 2. *In vitro* Callus induction response of different explant in EFY

Shoot Regeneration

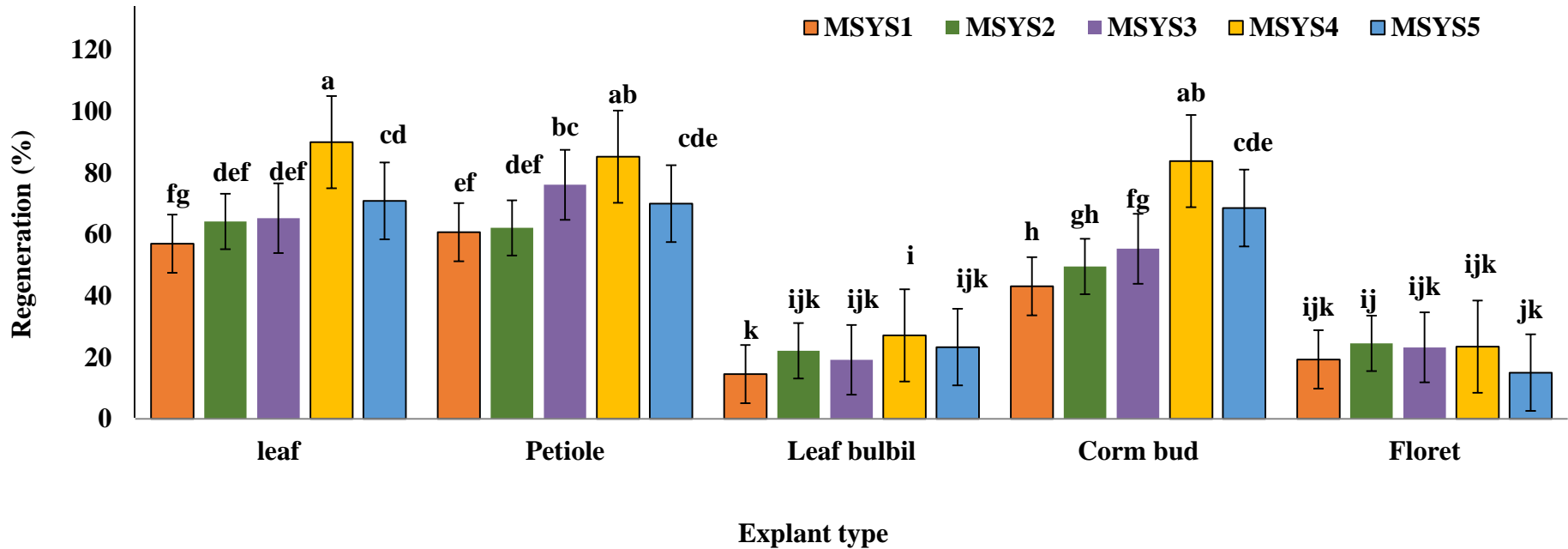


Figure 4. Effect of PGRs (BAP) on *in vitro* shoot regeneration of EFY from callus

*** a-k within a column having the same letters are not significantly different at 5% level

- Use of adenine sulphate in combination with cytokinins (BAP) achieve enhanced shoot multiplication from *in vitro* induced callus in potato. (Alok K.S *et al.*,2017)

Root Formation

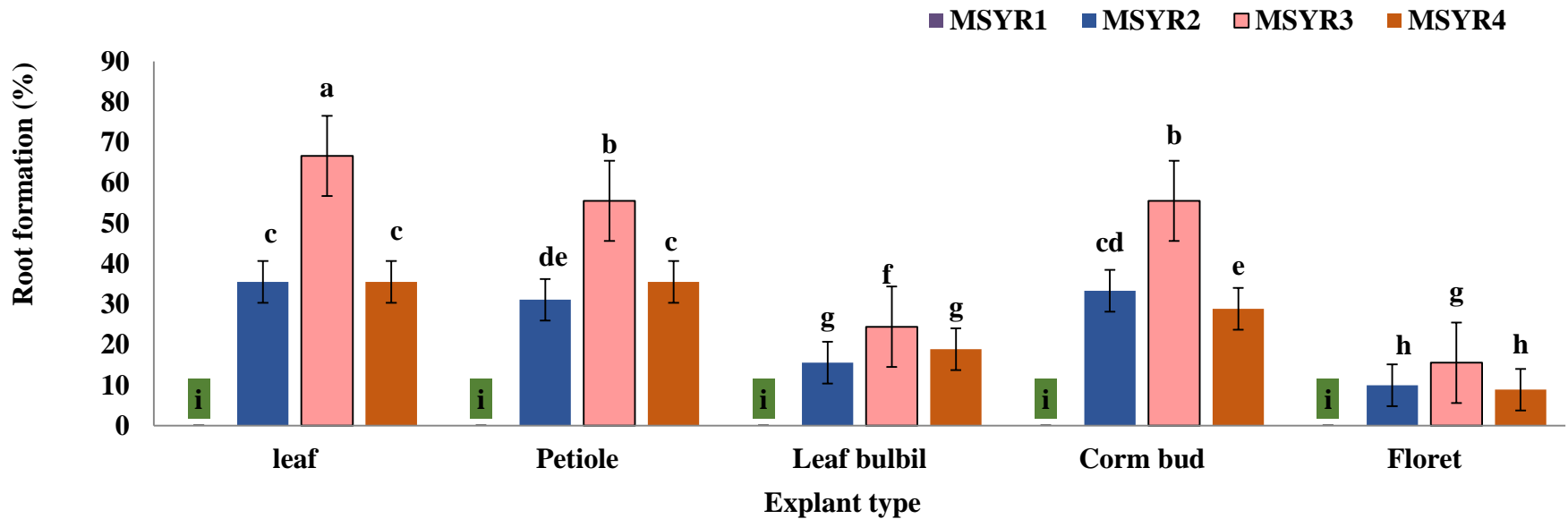


Figure 5. Effect of MS media and auxin (NAA) on rooting of EFY *in vitro* conditions

*** a-i within a column having the same letters are not significantly different at 5% level

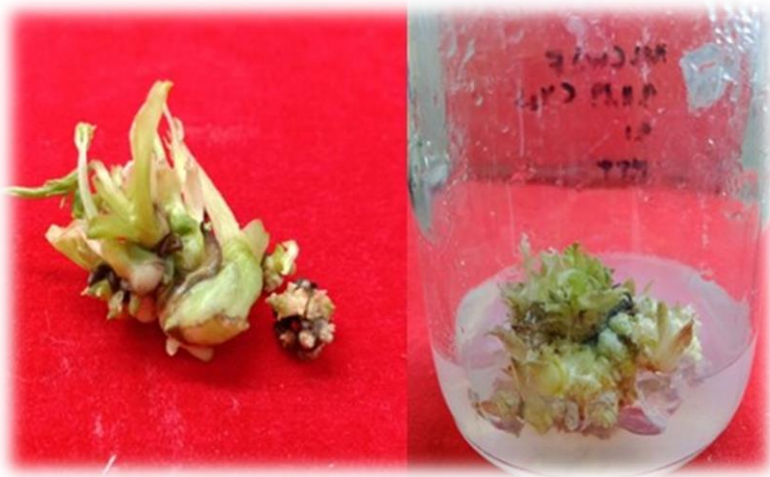
- *Amorphophallus konjac*, NAA proved to be efficient in root induction (Zhao et al., 2012), culturing shoots of *D.rotundata* (yam) on MS media with 0.5 mg/l NAA increase root formation (Sahar et al., 2017)

Table : Analysis of Variance for the effects of culture media and explants

Source (Callus Induction)	DF	MS	F value
Media	4	486.65	144.6**
Explants	4	333.74	99.16**
Media * Explant	16	29.27	8.70**
Error	48	3.37	
CV %	3.37		
LSD ^{0.05}	3.01		

Source (Shoot Regeneration)	DF	MS	F value
Media	4	495.43	29.44**
Explants	4	3670.8	218.12**
Media * Explant	16	65.12	3.87**
Error	48	16.83	
CV %	9.29		
LSD ^{0.05}	6.73		

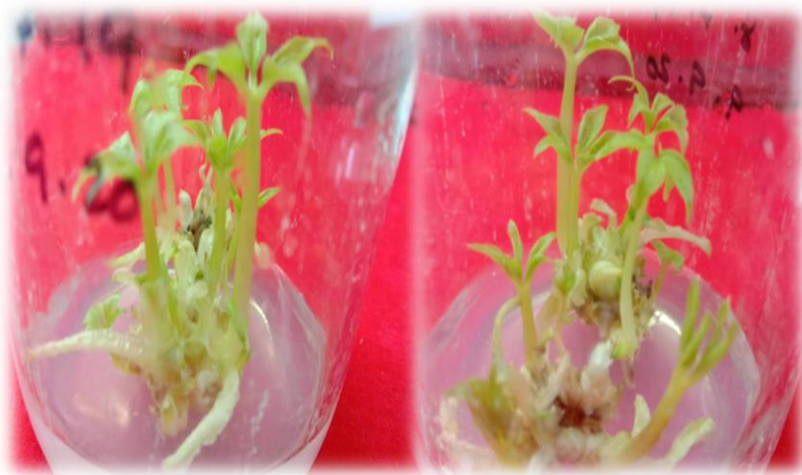
Source (Root Formation)	DF	MS	F value
Media	3	4582.21	1791.3**
Explants	4	663.9	259.53**
Media * Explant	12	98.44	38.48**
Error	38	2.56	
CV %	6.4		
LSD ^{0.05}	2.64		



(a) Shoot Induction



(b) Shoot Multiplication



(c) Shoot Elongation



(d) Root Formation

Figure 6. *In vitro* Shoot regeneration and root formation from Leaf explant of EFY

Hardening

- Remove well rooted plantlets and treat with 0.2% atonic for 5 min
- Transplant on potting mixture (soil, sand, burnt paddy husk, 1:1:1 in the net house)



Harden plant kept in the net house

Conclusions

Explant type

- Leaf is suitable explant material for production and preservation

Media combination

Callus induction

- **Modified MS + BAP (0.5 mg/l) + NAA (0.5mg/l) + 2,4-D (0.5mg/l)**

Shoot regeneration

- **Modified MS + BAP (4.0 mg/l) + Citric acid (1mg/l) + adenine sulphate (0.05g/l)**

Root formation

- **$\frac{1}{2}$ MS + NAA (1.0 mg/l)**

Future Trust

- Numerous plants will be produced all the year round and is more independent of seasonal changes
- The lost of plants can be minimized when transfer to natural environment and reduced production cost
- Providing the useful tools of tissue culture technology for mass propagation and conservation of Elephant Foot Yam

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